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Review Article A Comprehensive Review of Solvent-Induced Variability in Antioxidant Profiling of Plants Extract: *Justicia secunda*



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A B S T R A C T

Variability in antioxidant profiles, particularly within plant extracts like Justicia secunda and other plant extracts, presents a significant challenge for researchers striving to attain consistent and dependable outcomes in the realm of antioxidant profiling. This variability is predominantly attributed to the choice of solvents used during the extraction process. Our study delves deeply into the impact of solvent selection in the extraction process, revealing notable discrepancies in the antioxidant profiles of these plant extracts. This variation becomes evident through the diverse antioxidant potential observed across different solvents, emphasizing the pressing need for standardized methodologies to ensure research uniformity and reliability. Our review further explores the intricate interplay of diverse solvents employed during extraction procedures and their potential to induce variations in antioxidant profiles. It meticulously highlights the discernible fluctuations in antioxidant potential resulting from the use of different solvents, underscoring the imperative need for a systematic approach to research methodologies to guarantee consistent outcomes. The review concludes by presenting a forward-looking research agenda, including a comprehensive effort to identify and analyze specific antioxidant compounds within these extracts under various solvent conditions. Ultimately, our research seeks to enrich our understanding of the antioxidant properties and potential benefits associated with Justicia secunda. This review serves as an invaluable resource for researchers operating within this domain, spotlighting the pivotal role of solvent selection in antioxidant profiling and championing the cause of standardized methodologies to propel our knowledge forward in this area.

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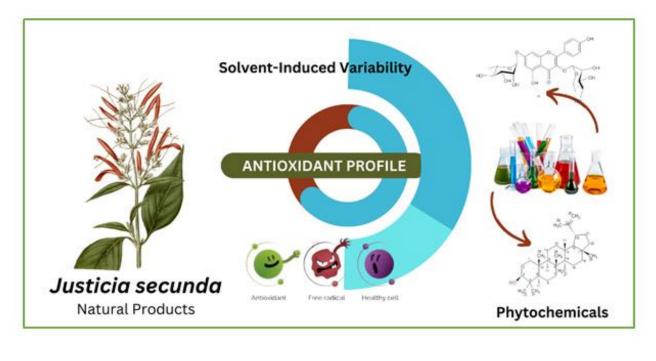
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GRAPHICAL ABSTRACT



Introduction

In recent years, the exploration of natural sources for bioactive compounds with potential health benefits has gained considerable attention in the scientific community. Plants, with their diverse array of phytochemicals, have become a primary focus of research, particularly in the pursuit of antioxidant compounds [1]. Antioxidants play a critical role in safeguarding the body against oxidative stress, a phenomenon linked to various chronic diseases, including cancer, cardiovascular disorders, and aging [3]. The study of antioxidants derived from plant sources, therefore, holds immense promise for the development of therapeutic and preventive strategies.

In the realm of natural compounds with potential health benefits, *Justicia secunda* stands as an intriguing subject of study. Its historical use in treating various ailments underscores its potential as a valuable resource in modern healthcare [1]. *Justicia secunda* is a medicinal plant widely distributed in tropical regions, and

has been traditionally used for its therapeutic properties. Previous research on Justicia secunda has reported its various pharmacological activities, including antioxidant, antiinflammatory, and analgesic effects [2]. The focus of this study was on the antioxidant profiles derived from its leaf and stem extracts, specifically exploring how different solvent extraction methods influence these profiles. Understanding the distribution of antioxidants in different parts of plants, such as leaves and stems, is essential. These plant components often possess unique phytochemical profiles, leading to varied antioxidant potential. In addition, the choice of solvents for extraction can significantly influence the types and quantities of antioxidants obtained [3].

The extraction of bioactive compounds from plant materials necessitates the use of solvents to separate and concentrate these compounds. It is this crucial stage of solvent selection that has

profound implications on the antioxidant potential of the resulting extracts. Our comprehensive review delves deep into this aspect, illuminating the remarkable influence that solvent choice wields over the antioxidant profiles of Justicia secunda extracts. We aimed to underline the urgency of implementing standardized methodologies to mitigate this variability, thus ensuring the uniformity and reliability of research outcomes.

This review serves as an invaluable resource, not only for researchers specifically interested in *Justicia secunda*, but also for the broader scientific community engaged in plant-based antioxidant profiling. Our research delves into the intricate interplay of diverse solvents used during the extraction procedures and the subsequent variations in antioxidant potential that they induce. By examining and comparing the diverse antioxidant potential observed across different solvents, we emphasize the paramount need for a systematic approach to research methodologies.

Furthermore, the implications of solvent-induced variability on the bioactivity of *Justicia secunda* extracts are discussed, shedding light on the potential consequences for applications within the realms of medicine and health. As these applications hold the promise of addressing pressing health issues, the need for reliable and consistent data becomes even more pronounced.

This exploration enhances our comprehension of the plant's overall antioxidant capacity. This review aims to provide a comprehensive comparative analysis of the antioxidant profiles in leaf and stem extracts of *Justicia secunda*. By employing various solvents for extraction, we seek to elucidate the influence of solvent variability on the observed antioxidant activity. This knowledge is pivotal for harnessing the full potential of *Justicia secunda* as a source of natural antioxidants with potential implications for human health.

Antioxidants and Their Significance

In our quest to unravel the intricacies of solventinduced variability in antioxidant profiling, we should initially acknowledge what antioxidants are and their profound importance in our review. These bioactive compounds (antioxidants) are the unsung heroes of our biological systems, silently combating the insidious effects of oxidative stress that assail our cells daily [22]. Oxidative stress, at its core, emerges from an imbalance between the production of harmful free radicals and the body's innate defense mechanisms [23-24]. Antioxidants emerge as the champions in this battle, swooping in to neutralize these free radicals and reactive oxygen species (ROS), preventing them from wreaking havoc on our cellular structures [25-27].

Relevance of Antioxidant Profiling

Antioxidants play a crucial role in protecting cells from oxidative damage caused by free radicals and reactive oxygen species (ROS) [28]. Antioxidant profiling is a valuable analytical tool in nutrition, medicine, and food science [29]. It involves the identification and quantification of various antioxidants present in a substance, such as food, plant extracts, or biological samples [30]. Profiling antioxidants in foods and dietary supplements helps assess their potential health benefits. This information aids consumers and healthcare professionals in making informed decisions about incorporating antioxidant-rich foods or supplements into their diets. Antioxidant profiling of foods can guide individuals in making dietary choices that promote health [31]. For example, people seeking to increase their intake of antioxidants can select foods known to be rich in specific antioxidants, like vitamin C, vitamin E, or flavonoids, based on profiling data.

Researchers use antioxidant profiling to study the antioxidant capacity of different foods, botanical extracts, and dietary patterns [32-33]. This information contributes to the understanding of

the role antioxidants play in preventing chronic diseases. such as heart disease, cancer, neurodegenerative disorders, and so on. In food and supplement industries, antioxidant profiling helps ensure product quality and consistency. Manufacturers can use these profiles to verify the presence and concentration of antioxidants claimed on product labels. This quality control process enhances consumer trust and safety [34]. In pharmaceutical research and development, antioxidant profiling can be vital in identifying potential drug candidates. Compounds with strong antioxidant properties may be explored for their therapeutic potential in treating diseases associated with oxidative stress [35-36].

Antioxidant profiling is useful in plant breeding and agriculture to develop crops with enhanced antioxidant content [37]. This can lead to the production of healthier foods with increased nutritional value. Antioxidant profiling of biological samples, such as blood or urine, can aid in identifying biomarkers of oxidative stress and related diseases [38]. These biomarkers can be used for diagnostic purposes and monitoring disease progression or treatment efficacy [39]. Antioxidant profiling can contribute to the emerging field of personalized nutrition. By assessing an individual's antioxidant status through profiling, tailored dietary and supplement recommendations can be made to address specific health needs [40]. Profiling antioxidants can also help monitor the effects of environmental pollutants, toxins, or contaminants on antioxidant levels in foods and ecosystems [41]. This information is essential for ensuring food safety and environmental health.

Health Benefits Associated with Antioxidants

Antioxidants are a cornerstone of holistic health, offering a profound array of benefits in combating chronic diseases and preserving well-being. From safeguarding cardiovascular health by reducing oxidative stress and inflammation to potentially thwarting cancer by protecting DNA integrity, their role is pivotal [42]. They hold promise in staving off neurodegenerative disorders, supporting the immune system, and nurturing skin health. By incorporating a diverse array of antioxidant-rich foods and embracing a healthy lifestyle, we harness the transformative potential of these compounds, equipping ourselves with a powerful defense against a range of health challenges and embracing the profound impact antioxidants have on our overall health and longevity.

Antioxidants play a crucial role in combating oxidative stress, a factor in various chronic diseases [16]. *Justicia secunda*, a plant with a rich history in traditional medicine, could hold valuable antioxidant compounds. Our research aims to uncover these compounds, paving the way for potential applications in health and wellness. Antioxidants, whether occurring naturally or synthesized, possess the potential to hinder or postpone specific forms of cellular harm [43]. These molecules combat free radicals within the body, these being substances that may inflict damage when their concentrations rise excessively [17]. Free radicals are perpetually generated within the body and can result in oxidative stress, a state that can impair cells and tissues, thereby contributing to chronic illnesses including cardiovascular diseases, diabetes, dementia, and cancer [18]. Antioxidants play a pivotal role in maintaining human health by counteracting oxidative stress, a process implicated in aging and various chronic diseases [19]. Natural antioxidants derived from plants, like those found in Justicia secunda, offer a promising avenue for promoting wellness and preventing oxidative damage [20].

Types of Antioxidants

Antioxidants are pivotal in protecting biological systems from the detrimental effects of oxidative stress by neutralizing harmful free radicals and reactive oxygen species (ROS). Their diverse forms constitute a multifaceted defense network against the relentless assault of free radicals and reactive oxygen species (ROS) [44]. This section delves into the various types of antioxidants, highlighting their distinct roles and significance in maintaining cellular health and overall wellbeing.

Enzymatic Antioxidants

Enzymatic antioxidants serve as the vanguard against oxidative stress. Superoxide dismutase (SOD) and catalase are exemplary defenders operating within cells [45]. SOD catalyzes the dismutation of superoxide radicals into hydrogen peroxide, while catalase subsequently breaks down hydrogen peroxide into harmless water and oxygen [46]. Together, these enzymes form the primary line of defense against superoxide and hydrogen peroxide, two highly reactive and damaging ROS. Their presence within cellular organelles ensures immediate neutralization of harmful radicals, preventing cellular damage and dysfunction [46].

Non-Enzymatic Antioxidants

Non-enzymatic antioxidants complement the enzymatic defense system, extending protection beyond the confines of cells [47,48]. Prominent among them are vitamins C and E, along with the versatile glutathione. Vitamin C, which is an aqueous-phase antioxidant, scavenges free radicals in body fluids and cellular cytoplasm, regenerating vitamin E in the process. Vitamin E, which is a lipid-soluble antioxidant, specializes in shielding cell membranes from lipid peroxidation [50]. Glutathione, often referred to as the "master antioxidant," orchestrates a cascade of reactions that detoxify harmful compounds and recycle other antioxidants, reinforcing the overall antioxidant defense system [51].

Phytochemical Antioxidants

Phytochemical antioxidants, which are found in abundance within plant extracts, comprise a

diverse range of compounds that make a significant contribution to our dietary intake of antioxidants. [52]. Flavonoids, carotenoids, and polyphenols are among the most studied phytochemical antioxidants. Flavonoids, found in fruits, vegetables, and beverages such as tea and red wine, possess diverse antioxidant properties, offering protection against oxidative damage [53]. Carotenoids which are responsible for the vibrant colors of many fruits and vegetables, are renowned for their ability to quench singlet oxygen and protect against photooxidative processes [53, 55]. Polyphenols found in foods like berries, nuts, and dark chocolates, exhibit antioxidant, anti-inflammatory, and potential anti-cancer properties [54].

Overview of Methods and Solvents Used for Antioxidant Profile from J. secunda Extracts and other plants

Assessing the antioxidant properties of plant extracts is crucial for understanding their potential health benefits. However, the use of different extraction methods and solvents in various studies can make it challenging to compare and draw meaningful conclusions about the antioxidant profiles of "Justicia secunda" (J. secunda) and other plants. Many researchers have caried similar studies on the antioxidant profile using different solvents and methods, as presented in Table 1.

Overview of Phytochemicals Composition of Justicia secunda and Its Potential Bioactive (Antioxidant) Compounds

Phytochemicals are bioactive compounds that occur naturally in plants. They contribute to the plants' color, taste, and smell, while also serving essential functions in defending the plant and interacting with its environment [4]. *Justicia secunda*, a plant of interest in botanical and pharmaceutical research, has been a subject of investigation due to its potential therapeutic properties attributed to its phytochemical content [102]. *Justicia secunda* is rich in a diverse array of bioactive compounds, including flavonoids, alkaloids, terpenoids, phenolics, and glycosides. These compounds, abundant in both its leaves and stems, are responsible for the plant's medicinal properties. Flavonoids, for instance, exhibit antioxidant, anti-inflammatory, and anticancer activities. Alkaloids may possess analgesic and antimicrobial properties, while terpenoids contribute to various therapeutic effects [5]. This botanical species offers a captivating case study of how distinct plant parts can harbor unique assortments of bioactive compounds, reflecting

their specialized functions within the plant's lifecycle. These distinctive phytochemical profiles are orchestrated by a harmonious interplay of intrinsic physiological roles and extrinsic environmental cues. In this species, the leaves and stems exhibit noteworthy variations in their phytochemical profiles, with each part containing a unique array of compounds that serve diverse biological functions. These variations are the result of intricate biochemical processes influenced by both intrinsic factors related to plant physiology and extrinsic factors like environmental conditions [5].

Table 1.Comparison of Extraction Methods and Solvent Used for Antioxidant Profile from J. secunda Extracts andother plants

Plants	Extraction Method	Solvent Used	Antioxidant Activity	Ref.
<i>Justicia secunda</i> Leaf and Stem		Hexane	Highest	[96]
		Ethyl acetate	Significant	
		Acetone	Significant	
	Cold Maceration	Methanol	Significant	
Justicia secunda		Dichloromet hane	Significant	[<u>21]</u>
		Ethyl acetate	Significant	
		Methanol	Highest	
	Maceration and Infusion	Water	Highest	
Dipsacus asperoides		Acetone	Significant	[100]
	Ultrasonic-assisted Extraction with Filtration and	Methanol,	Highest	
	Concentration.	Water	Significant	
Justicia secunda Vahl Leaf	Maceration	Methanol	High	[<u>8]</u>
Justicia secunda Vahl	Maceration	Methanol	High	[99]

Significant variations in phytochemical composition exist between the leaves and stems of *Justicia secunda*. Studies have shown that leaves may contain higher concentrations of certain flavonoids, such as quercetin, whereas stems may exhibit elevated levels of alkaloids like vincamine [56]. These differences in specific compounds could be attributed to distinct physiological functions of leaves and stems within the plant, as well as environmental factors [6].

The distinct phytochemical profiles of *Justicia secunda*'s leaves and stems hold promising implications for potential health benefits [97]. Utilizing different parts of the plant in traditional medicine or herbal remedies may target specific health conditions [7,8].

For example, the higher flavonoid content in the leaves may make them more suitable for antioxidant and anti-inflammatory applications, while the stem extracts, rich in alkaloids, could be valuable for their analgesic or antimicrobial effects.

Understanding these differences allows for targeted utilization of *Justicia secunda*'s various parts to maximize its therapeutic potential and contribute to the development of natural health products.

Role of Specific Bioactive Compounds in Contributing to Antioxidant Activity Differences among Solvents

The presence of unique bioactive compounds in Justicia secunda plays a crucial role in the observed differences in antioxidant activity among solvents. Some solvents may be more efficient at extracting specific compounds due to their chemical nature [96]. For instance, certain solvents might excel at extracting flavonoids, while others are better suited for alkaloids. Understanding the distribution of these compounds in different solvents provides insights into the targeted extraction of specific antioxidants and the potential modulation of *Justicia secunda*'s bioactivity based on solvent selection.

The interplay between solvent properties, extraction conditions, and the specific bioactive compounds present in *Justicia secunda* contributes to the variability in antioxidant profiles [98].

This knowledge enables researchers to strategically select solvents for extracting antioxidants of interest, ensuring that the unique health-promoting properties of different bioactive compounds are effectively harnessed for potential therapeutic applications.

Solvent Extraction Methods

Solvent extraction is a commonly utilized method for extracting bioactive compounds from plant materials and involves using various techniques to isolate these compounds, including maceration, Soxhlet extraction, and ultrasound-assisted extraction [57].

Maceration

Maceration is a conventional approach to solvent extraction, employed for the isolation of bioactive substances from botanical materials [58-60]. In this method, plant materials are submerged or soaked in a suitable solvent like hexane, ethanol, or acetone for an extended period, often spanning from several days to weeks. During this period, the solvent gradually permeates the plant material, causing the dissolution of the desired bioactive components. Maceration is known for its simplicity and cost-effectiveness, making it a favored choice for extracting compounds such as phytochemicals, phenolics, and alkaloids. However, it is important to note that it may necessitate a longer extraction duration when compared to more modern methodologies [57].

Soxhlet Extraction

Soxhlet extraction is another widely embraced technique for the extraction of bioactive

compounds from plant sources. This method is particularly effective for extracting compounds with limited solubility in conventional solvents. It entails a continuous cycle of extraction, reflux, and condensation [61]. The plant material is typically placed within a porous thimble, which is then inserted into a Soxhlet extractor. A solvent, often a high-boiling-point solvent such as hexane, is heated, vaporized, and subsequently condensed back into the extraction chamber, effectively cycling the solvent through the plant material. This process continues until the extracted compounds are concentrated within the solvent. Soxhlet extraction is renowned for its efficiency and its capability to extract a diverse array of compounds [62].

Ultrasound-Assisted Extraction (UAE)

Ultrasound-assisted extraction is a contemporary and highly efficient method that harnesses ultrasound waves to augment the extraction process. In this technique, the plant material is mixed with a suitable solvent within an ultrasound bath [63]. The application of ultrasound energy generates cavitation bubbles within the solvent, which implode in proximity to the plant material's surface. This phenomenon creates localized microenvironments characterized by elevated temperatures and pressures, thereby facilitating the extraction of bioactive substances. UAE is applauded for its rapid extraction speed, reduced solvent consumption, and enhanced yield of target compounds. It is often the preferred choice for extracting heat-sensitive substances such as specific polyphenols and essential oils [64].

Each of these solvent extraction methods offers unique advantages, with the selection contingent upon the specific characteristics of the plant material and the targeted bioactive compounds [65]. Researchers judiciously opt for the most appropriate technique to optimize the extraction process and maximize the yield of bioactive constituents for subsequent analysis and potential applications in diverse fields, including pharmaceuticals, food production, and cosmetics. The choice of technique depends on the type of plant material, the desired compounds, and the equipment available. Maceration involves soaking the plant material in a solvent to allow for compound dissolution, while Soxhlet extraction employs continuous solvent cycling. The ultrasound-assisted extraction uses ultrasonic waves to enhance compound release from the plant matrix, facilitating higher extraction efficiency [9].

Among the methods mentioned for isolating bioactive compounds from plant materials, the ultrasound-assisted extraction (UAE) has gained significant attention and popularity among researchers in recent years. This technique has shown promising results in terms of higher extraction efficiency and reduced extraction time compared to traditional methods like maceration and Soxhlet extraction [66]. Researchers have utilized ultrasound-assisted extraction in various studies to extract bioactive compounds from plant materials. [10] investigated the use of ultrasoundassisted extraction for obtaining bioactive compounds from plants. They found that the UAE significantly improved the extraction efficiency and reduced the extraction time compared to traditional methods. In a study by [11] reviewed the use of ultrasound-assisted extraction for plant bioactive compounds, including phenolics, flavonoids, thymols, saponins, and proteins. The concluded that ultrasound-assisted review extraction is a green extraction technique that offers high performance with less solvent and time consumption. The study also found that ultrasound-assisted extraction can extract bioactive components in less time, at low temperature, with lesser energy and solvent consumption. The study suggests that ultrasoundassisted extraction is an effective method for extracting bioactive compounds from plants. Another study by [12] focused on extracting polyphenols from Lonicera japonica (Japanese honeysuckle) flowers using ultrasound-assisted extraction. They found that this method significantly improved the extraction efficiency and enabled the recovery of a higher amount of polyphenolic compounds.

Overall, ultrasound-assisted extraction has demonstrated its effectiveness in enhancing the extraction of bioactive compounds from plant materials. Researchers have reported improved extraction efficiency, shorter extraction times, and higher yields of target compounds. However, the success of the technique can also depend on factors such as the type of plant material, the solvent used, and the specific bioactive compounds of interest. It is important to note that while the UAE has shown promise, the choice of extraction method may still vary based on the research goals and the characteristics of the plant material being studied.

Factors Influencing Solvent Selection for Antioxidant Extraction

The choice of solvent in antioxidant profiling plays a pivotal role in the effectiveness of the extraction process and the quality of obtained results [67]. Researchers should carefully consider several factors when selecting the most suitable solvent for their specific study. These factors need to be carefully considered to ensure accurate and effective extraction of bioactive compounds. According to [3], factors such as solvent polarity, selectivity, and compatibility with the target compounds influence the choice. The nature of the bioactive compounds in Justicia secunda, as well as their desired application, dictates the suitable solvent. The solvent polarity impacts the extraction of specific phytochemicals, with polar solvents being more effective for extracting polar compounds like flavonoids, while non-polar solvents may better extract lipophilic compounds [13]. Here are the key factors that influence solvent selection in antioxidant profiling:

Solubility of Target Compounds

The solubility of antioxidants in a chosen solvent is a fundamental factor. Different antioxidants exhibit varying degrees of solubility based on their chemical properties [68]. For example, hydrophilic antioxidants, such as ascorbic acid or polyphenols, are more soluble in polar solvents like water, methanol, or ethanol. In contrast, lipophilic antioxidants, including carotenoids or tocopherols, tend to dissolve better in nonpolar solvents like hexane or ethyl acetate [69]. Therefore, carefully assessment of the chemical characteristics of their target antioxidants to select a solvent that facilitates efficient extraction.

Nature of Plant Material

The composition of the plant material under investigation significantly influences solvent selection [96]. Different plant tissues contain a wide range of phytochemicals, and the choice of solvent should align with the constituents' present [98]. For example, extracting antioxidants from fruit peels, which often contain lipophilic compounds, may necessitate the use of nonpolar solvents. Conversely, extracting antioxidants from leaves or stems, which may contain hydrophilic compounds, may require polar solvents [70].

Heat Sensitivity

Some antioxidants are sensitive to heat, and exposure to elevated temperatures during the extraction process can lead to degradation and loss of bioactivity. To preserve the integrity of these compounds, researchers opt for solvents that allow for extraction at lower temperatures. Cold solvents, such as cold methanol or ethanol, are often chosen to prevent heat-induced degradation while still achieving effective extraction [71].

Safety and Toxicity

Safety considerations are paramount in solvent selection. Some solvents, such as chloroform or

dichloromethane, may pose health risks or have adverse environmental impacts, and thus should be used with caution or avoided [72]. The safety data sheets for solvents provide essential information about their toxicity, flammability, and safe handling procedures [73].

Cost and Availability

It is noteworthy that practical factors like cost and availability influence solvent selection. Certain solvents may be expensive, particularly those of high purity grades, and could significantly impact research budgets. In addition, the availability of solvents in the required quantities and grades can be a logistical consideration, particularly for large-scale studies [74].

Extraction Efficiency

The efficiency of a solvent in extracting antioxidants is a crucial factor. Researchers aim to achieve high extraction yields while minimizing solvent usage. Solvents that efficiently extract target compounds without excessive waste are preferred to optimize resource utilization and minimize environmental impact [75].

Compatibility with Analytical Techniques

Solvent compatibility with the chosen analytical techniques is essential for obtaining accurate and reliable results. Different analytical methods, such as spectrophotometry, high-performance liquid chromatography (HPLC), or gas chromatography (GC), may require specific solvents to ensure proper sample preparation and reliable measurements. Compatibility considerations include the solvent's chemical compatibility with the analytical instrument and its ability to provide clean and interference-free extracts [76].

Regulatory Compliance

Regulatory compliance is vital, especially in applications related to food, pharmaceuticals, or cosmetics [77]. Researchers should adhere to regulatory guidelines and standards when working with solvents to ensure the safety and quality of the final products. Compliance includes following good laboratory practices, handling hazardous materials in accordance with safety regulations, and ensuring that solvents used meet regulatory purity standards.

Impact of Solvent Choice on Antioxidant Profiling

Several solvents are commonly employed in plant extraction, each with its advantages and limitations. Several studies have explored the variability in antioxidant profiling of *Justicia Secunda* extracts resulting from various solvent choices. Notably, ethanol, methanol, acetone, and water are among the commonly employed solvents [15,98].

1. Ethanol Extracts: Ethanol is frequently used for extracting antioxidants from Justicia secunda [78]. Research shows that ethanol-based extractions yield a diverse range of antioxidants, including phenolic compounds and flavonoids. These extracts have demonstrated strong free radical scavenging activity and significant potential for health-promoting properties [79].

2. Methanol Extracts: Methanol is another widely used solvent for plants [81]. Studies suggest that methanol extractions are effective at capturing a broad spectrum of antioxidants, particularly polyphenols [80]. Methanol-based extracts exhibit substantial antioxidant activity and are valued for their potential in combating oxidative stress-related diseases [82].

3. Acetone Extracts: Acetone is chosen in some studies for its ability to extract antioxidants efficiently. Acetone-based extracts of *Justicia secunda* have been found to contain various polyphenols and flavonoids. These extracts exhibit antioxidant potential, contributing to the plant's overall therapeutic value [82].

4. Water Extracts: Water extraction, while milder than organic solvents, remains a valuable method for obtaining antioxidants from *Justicia secunda* [83]. Water-based extracts are rich in water-

soluble antioxidants, such as vitamin C, and exhibit antioxidant activity, albeit to a lesser extent than organic solvent extracts [84].

Polar solvents like methanol and ethanol offer high extraction efficiency for a broad range of compounds, but might also co-extract undesired substances. Non-polar solvents such as hexane are effective for lipophilic compounds but may miss some polar antioxidants. Ethyl acetate combines characteristics of both polar and nonpolar solvents, making it suitable for diverse phytochemicals, yet its use requires careful consideration due to potential toxicity [14].

Methodologies in Antioxidant Profiling

Antioxidant profiling relies on a range of analytical techniques, each with its strengths and applications [20]. The choice of technique should align with research objectives, sample type, and the specific antioxidants of interest. Furthermore, sample preparation techniques play a crucial role in minimizing variability and ensuring the accuracy of antioxidant analysis. This review discusses the diverse methodologies and analytical techniques commonly employed in antioxidant profiling, emphasizing their importance in elucidating the antioxidant capacity of various substances. In addition, we explore the relevance of sample preparation techniques and their impact on solvent-induced variability.

1. Spectrophotometry: Spectrophotometry is a widely used technique for assessing antioxidant activity based on the absorbance measurement at specific wavelengths. The most common assays include the DPPH (2,2-diphenyl-1-picrylhydrazyl) and ABTS (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)) assays. Spectrophotometric assays are rapid and cost-effective, providing a preliminary assessment of antioxidant capacity [85].

2. HPLC (High-Performance Liquid Chromatography): HPLC is a powerful tool for identifying and quantifying specific antioxidants within complex mixtures. It enables the separation of compounds based on their chemical properties, such as polarity and molecular weight. HPLC coupled with various detectors, such as UV-Vis, fluorescence, or mass spectrometry, allows for precise quantification of antioxidants, including vitamins (e.g., vitamin C and E) and polyphenols [86].

3. Gas Chromatography (GC): GC is primarily used for analyzing volatile antioxidants and lipidsoluble antioxidants. It involves the vaporization of compounds and their separation based on their volatility. GC is especially valuable for studying antioxidants like carotenoids and tocopherols, commonly found in lipids [87].

4. Liquid Chromatography-Mass Spectrometry (LC-MS): LC-MS combines the separation capabilities of liquid chromatography with mass spectrometry's specificity, making it suitable for identifying and quantifying antioxidants. LC-MS can handle complex samples and is essential for characterizing the structure of novel antioxidants [88].

In the context of Justicia secunda, various antioxidant assays have been employed to assess the antioxidant activity of its leaf and stem extracts. These assays were chosen based on their relevance to the specific bioactive compounds present in Justicia secunda and their ability to provide comprehensive information on antioxidant potential. The rationale for selecting assays involves their sensitivity, these reproducibility, and established use in similar plant studies [21]. The influence of solvent variability on the observed antioxidant activity is discussed, with a focus on the unique antioxidant capacities of each plant part. The combination of selected antioxidant assays, the unique phytochemical composition of Justicia secunda, and the use of different solvents allows for a comprehensive evaluation of the antioxidant potential of the plant's extracts. This information is valuable for understanding the role of Justicia secunda in promoting health and its potential applications in the development of natural antioxidant-based products.

Implications of Solvent-Induced Variability

Solvent choice in antioxidant profiling significantly impacts the composition and potency of plant extracts. This variability has several implications, as follow [89]:

Health Applications

Given the potential health benefits of *Justicia secunda* antioxidants, such as cardiovascular protection and neuroprotection, solvent-induced variability can influence the efficacy of herbal remedies and dietary supplements. Inconsistent antioxidant profiles may lead to variations in therapeutic outcomes, posing challenges for healthcare practitioners and consumers seeking standardized treatments [90].

Pharmaceutical Industry

The pharmaceutical sector is increasingly exploring the therapeutic potential of natural antioxidants. Solvent-induced variability raises concerns about the consistency and efficacy of pharmaceutical products derived from *Justicia secunda*. Standardization becomes crucial to ensure reliable and reproducible formulations [91].

Food Industry

Justicia secunda is utilized in traditional foods and functional beverages [103]. Variability in antioxidant content can impact product quality and labeling accuracy. Food manufacturers need to address this variability to make credible health claims and meet consumer expectations [92].

Biomedical Research

Researchers studying plant's antioxidant properties should consider solvent-induced variability when designing experiments and interpreting results because the choice of solvent can significantly impact the extraction efficiency and, consequently, the antioxidant activity of phytochemicals from the plant material. Inaccurate or inconsistent antioxidant profiling can lead to erroneous conclusions about its health benefits and potential mechanisms of action [93].

Challenges and Opportunities for Standardization

Standardization of antioxidant profiling methodologies is essential to address solventinduced variability and unlock the full potential of plant's extracts [94]. Some of the challenges and opportunities include:

Methodological Diversity

The plethora of analytical techniques and solvents used in antioxidant profiling poses a challenge to standardization [94]. Collaborative efforts among researchers, institutions, and regulatory bodies are needed to develop standardized protocols.

Reference Materials

The availability of reference materials, including certified antioxidant standards, can aid in method validation and comparison [94]. Developing a comprehensive database of reference materials specific to *Justicia secunda* can enhance standardization efforts.

Regulatory Guidelines

Regulatory bodies should establish clear guidelines for antioxidant profiling in herbal products and supplements. These guidelines can define acceptable methods, reporting standards, and acceptable levels of variability [95].

Data Sharing

Encouraging researchers to openly share their data and methodologies can foster collaboration and drive standardization efforts. Transparent reporting of extraction protocols, used solvents, and antioxidant quantification methods is critical.

Solvent-induced variability in antioxidant profiling of plant extracts has far-reaching implications for health and industrial applications. Addressing these challenges through standardization efforts is paramount to ensure the reliability and reproducibility of results, ultimately benefiting consumers, the pharmaceutical industry, food manufacturers, and biomedical research. Standardization will not only enhance the credibility of plants therapeutic potential, but also contribute to the broader field of antioxidant research, fostering trust in natural remedies and products.

Conclusion

This entitled: "A comprehensive review Comprehensive Review of Solvent-Induced Variability in Antioxidant Profiling of Plants Extract: Justicia secunda" emphasizes the critical role of solvent selection in antioxidant profiling, particularly concerning Justicia secunda. A key takeaway is the significant impact that different solvents can have on the assessment of antioxidant activity in Justicia secunda extracts. It underscores the need for researchers to exercise careful consideration when choosing solvents, as this choice profoundly influences the reliability and outcomes of their studies. Moreover, the review highlights the substantial variability in antioxidant potential observed across various solvents, with some yielding higher concentrations while others yield lower ones. This inherent variability necessitates a nuanced approach to study Justicia secunda's antioxidant properties. To address these challenges, future research should focus on identifying specific antioxidant compounds within Justicia secunda extracts under different solvent conditions and standardizing extraction and analysis methods to enhance consistency and reliability in antioxidant profiling, ultimately advancing our understanding of this plant's antioxidant potential and its broader implications.

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References

- Bian T, Corral P, Wang Y, Botello J, Kingston R, Daniels T, Salloum RG, Johnston E, Huo Z, Lu J, Liu AC. Kava as a clinical nutrient: promises and challenges, Nutrients; 2020 Oct 5; 12(10):3044. [Crossref], [Google Scholar], [Publisher]
- 2. SRIDhaR N, Duggirala SL, Puchchakayala G. Evaluation of Antioxidant and Anti-

Inflammatory Activities of Justicia neesii Ramam Whole Plant Extract. Journal of Pharmaceutical Technology, Research and Management; 2014 Nov 30; 2(2):171-87. [Crossref], [Google Scholar], [Publisher]

- 3. Boeing JS, Barizão ÉO, e Silva BC, Montanher PF, de Cinque Almeida V, Visentainer JV. Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis, Chemistry central journal; 2014 Dec; 8:1-9. [Crossref], [Google Scholar], [Publisher]
- Rabizadeh F, Mirian MS, Doosti R, Kiani-Anbouhi R, Eftekhari E. Phytochemical Classification of Medicinal Plants Used in the Treatment of Kidney Disease Based on Traditional Persian Medicine, Evidence-Based Complementary and Alternative Medicine; 2022 Jul 31; 2022. [Crossref], [Google Scholar], [Publisher]
- 5. Theiler BA, Revoltella S, Zehl M, Dangl C, Caisa LO, König J, Winkler J, Urban E, Glasl S. Secundarellone A, B, and C from the leaves of Justicia secunda Vahl, Phytochemistry Letters; 2014 Dec 1; 10:cxxix-xii. [Crossref], [Google Scholar], [Publisher]
- 6. Roy A, Khan A, Ahmad I, Alghamdi S, Rajab BS, Babalghith AO, Alshahrani MY, Islam S, Islam MR. Flavonoids a bioactive compound from medicinal plants and its therapeutic applications, BioMed Research International; 2022 Jun 6; 2022. [Crossref], [Google Scholar], [Publisher]
- Anyasor GN, Okanlawon AA, Ogunbiyi B. Evaluation of anti-inflammatory activity of Justicia secunda Vahl leaf extract using in vitro and in vivo inflammation models, Clinical Phytoscience; 2019 Dec; 5:1-3. [Crossref], [Google Scholar], [Publisher]
- 8. Onoja SO, Ezeja MI, Omeh YN, Onwukwe BC. Antioxidant, anti-inflammatory and antinociceptive activities of methanolic extract of Justicia secunda Vahl leaf,

Alexandria Journal of Medicine; 2017 Dec 6; 53(3):207-13. [Crossref], [Google Scholar], [Publisher]

- Bitwell C, Sen IS, Luke C, Kakoma MK. A review of modern and conventional extraction techniques and their applications for extracting phytochemicals from plants. Scientific African. 2023 Feb 11:e01585. [Crossref], [Google Scholar], [Publisher]
- Yusoff IM, Taher ZM, Rahmat Z, Chua LS. A review of ultrasound-assisted extraction for plant bioactive compounds: Phenolics, flavonoids, thymols, saponins and proteins. Food research international. 2022 Jul 1;157:111268. [Crossref], [Google Scholar], [Publisher]
- 11. Yusoff IM, Taher ZM, Rahmat Z, Chua LS. A review of ultrasound-assisted extraction for plant bioactive compounds: Phenolics, flavonoids, thymols, saponins and proteins, Food research international; 2022 Jul 1; 157:111268. [Crossref], [Google Scholar], [Publisher]
- 12. DIMITRIU L, PREDA D, CONSTANTINESCU-ARUXANDEI D, OANCEA F, BĂBEANU N. Optimization of ultrasound-assisted extraction of polyphenols from honeysuckle (Lonicera caprifolium), AgroLife Scientific Journal; 2021 Dec 15; 10(2). [Crossref], [Google Scholar], [Publisher]
- 13. Nawaz H, Shad MA, Rehman N, Andaleeb H, Ullah N. Effect of solvent polarity on extraction yield and antioxidant properties of phytochemicals from bean (Phaseolus vulgaris) seeds, Brazilian Journal of Pharmaceutical Sciences; 2020 Mar 16; 56:e17129. [Crossref], [Google Scholar], [Publisher]
- Altemimi A, Lakhssassi N, Baharlouei A, Watson DG, Lightfoot DA. Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts, Plants; 2017 Sep 22; 6(4):42. [Crossref], [Google Scholar], [Publisher]

- 15. Chemat F, Abert Vian M, Ravi HK, Khadhraoui B, Hilali S, Perino S, Fabiano Tixier AS. Review of alternative solvents for green extraction of food and natural products: Panorama, principles, applications and prospects, Molecules; 2019 Aug 19; 24(16):3007. [Crossref], [Google Scholar], [Publisher]
- Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health, Pharmacognosy reviews; 2010 Jul; 4(8):118. [Crossref], [Google Scholar], [Publisher]
- 17. Arnarson, A. (2023, July 12). Antioxidants Explained in Simple Terms. Healthline. https://www.healthline.com/nutr ition/antioxidants-explained. [CrossRef], [Google Scholar], [Publisher]
- WebMD Editorial Contributors. (2023, April 27). What Is Oxidative Stress? Retrieved from https://www.webmd.com/a-to-zguides/what-is-oxidative-stress. [CrossRef], [Google Scholar], [Publisher]
- Forman HJ, Zhang H. Targeting oxidative stress in disease: Promise and limitations of antioxidant therapy, Nature Reviews Drug Discovery; 2021 Sep; 20(9):689-709. [Crossref], [Google Scholar], [Publisher]
- 20. Chaves N, Santiago A, Alías JC. Quantification of the antioxidant activity of plant extracts: Analysis of sensitivity and hierarchization based on the method used, Antioxidants; 2020 Jan 15; 9(1):76. [Crossref], [Google Scholar], [Publisher]
- 21. Świątek Ł, Sieniawska E, Sinan KI, Zengin G, Boguszewska A, Hryć B, Bene K, Polz-Dacewicz M, Dall'Acqua S. Chemical Characterization of Different Extracts of Justicia secunda Vahl and Determination of Their Anti-Oxidant, Anti-Enzymatic, Anti-Viral, and Cytotoxic Properties, Antioxidants; 2023 Feb 17; 12(2):509. [Crossref], [Google Scholar], [Publisher]
- 22. Singh R, Devi S, Gollen R. Role of free radical in atherosclerosis, diabetes and

dyslipidaemia: larger-than-life, Diabetes/metabolism research and reviews; 2015 Feb; 31(2):113-26. [Crossref], [Google Scholar], [Publisher]

- Shah MN. The role of free radicals and reactive oxygen species in biological systemsa comprehensive review, International Journal Of Drug Research And Dental Science; 2022 Oct 5; 4(3):28-41. [Crossref], [Google Scholar], [Publisher]
- 24. Sundaram Sanjay S, Shukla AK. Free radicals versus antioxidants. InPotential Therapeutic Applications of Nano-antioxidants 2021 Aug 28 (pp. 1-17). Singapore: Springer Singapore. [Crossref], [Google Scholar], [Publisher]
- 25. Manchester LC, Coto-Montes A, Boga JA, Andersen LP, Zhou Z, Galano A, Vriend J, Tan DX, Reiter RJ. Melatonin: an ancient molecule that makes oxygen metabolically tolerable, Journal of pineal research; 2015 Nov; 59(4):403-19. [Crossref], [Google Scholar], [Publisher]
- 26. Adwas AA, Elsayed A, Azab AE, Quwaydir FA. Oxidative stress and antioxidant mechanisms in human body, J. Appl. Biotechnol. Bioeng; 2019 Feb 21; 6(1):43-7. [Crossref], [Google Scholar], [Publisher]
- 27. Szeto YT, Tomlinson B, Benzie IF. Total antioxidant and ascorbic acid content of fresh fruits and vegetables: implications for dietary planning and food preservation, British journal of nutrition; 2002 Jan; 87(1):55-9. [Crossref], [Google Scholar], [Publisher]
- Oroian M, Escriche I. Antioxidants: Characterization, natural sources, extraction and analysis, Food Research International; 2015 Aug 1; 74:10-36. [Crossref], [Google Scholar], [Publisher]
- 29. Halliwell B. Free radicals and antioxidants: updating a personal view, Nutrition reviews; 2012 May 1; 70(5):257-65. [Crossref], [Google Scholar], [Publisher]
- 30. Butnariu M, Coradini CZ. Evaluation of biologically active compounds from Calendula

officinalis flowers using spectrophotometry, Chemistry central journal; 2012 Dec; 6:1-7. [Crossref], [Google Scholar], [Publisher]

- Benzie IF, Choi SW. Antioxidants in food: content, measurement, significance, action, cautions, caveats, and research needs, *Advances in food and nutrition research*; 2014 Jan 1; 71:1-53. [CrossRef], [Google Scholar], [Publisher]
- 32. Pellegrini N, Vitaglione P, Granato D, Fogliano V. Twenty-five years of total antioxidant capacity measurement of foods and biological fluids: merits and limitations, *Journal of the Science of Food and Agriculture*; 2020 Nov; 100(14):5064-78. [CrossRef], [Google Scholar], [Publisher]
- 33. Wang S, Meckling KA, Marcone MF, Kakuda Y, Tsao R. Synergistic, additive, and antagonistic effects of food mixtures on total antioxidant capacities, *Journal of agricultural and food chemistry*; 2011 Feb 9; 59(3):960-8. [CrossRef], [Google Scholar], [Publisher]
- Ravichandran 34. R. Nanotechnology applications in food and food processing: innovative green approaches, opportunities uncertainties for global and market, International **Journal** of Green Nanotechnology: Physics and Chemistry; 2010 May 20; 1(2):P72-96. [CrossRef], [Google Scholar], [Publisher]
- 35. White PA, Oliveira RC, Oliveira AP, Serafini MR, Araújo AA, Gelain DP, Moreira JC, Almeida JR, Quintans JS, Quintans-Junior LJ, Santos MR. Antioxidant activity and mechanisms of action of natural compounds isolated from lichens: A systematic review, Molecules; 2014 Sep 12; 19(9):14496-527. [CrossRef], [Google Scholar], [Publisher]
- 36. Ratnam DV, Ankola DD, Bhardwaj V, Sahana DK, Kumar MR. Role of antioxidants in prophylaxis and therapy: A pharmaceutical perspective, *Journal of controlled release*; 2006 Jul 20; 113(3):189-207. [CrossRef], [Google Scholar], [Publisher]

- 37. Frary A, Göl D, Keleş D, Ökmen B, Pınar H, Şığva HÖ, Yemenicioğlu A, Doğanlar S. Salt tolerance in Solanum pennellii: antioxidant response and related QTL, *BMC Plant Biology*; 2010 Dec; 10(1):1-6. [CrossRef], [Google Scholar], [Publisher]
- 38. Ho E, Galougahi KK, Liu CC, Bhindi R, Figtree GA. Biological markers of oxidative stress: applications to cardiovascular research and practice, *Redox biology*; 2013 Jan 1; 1(1):483-91. [CrossRef], [Google Scholar], [Publisher]
- 39. Mueller SG, Weiner MW, Thal LJ, Petersen RC, Jack CR, Jagust W, Trojanowski JQ, Toga AW, Beckett L. Ways toward an early diagnosis in Alzheimer's disease: the Alzheimer's Disease Neuroimaging Initiative (ADNI), *Alzheimer's & Dementia*; 2005 Jul 1;1(1):55-66. [CrossRef], [Google Scholar], [Publisher]
- 40. Margaritelis NV, Paschalis V, Theodorou AA, Kyparos A, Nikolaidis MG. Antioxidants in personalized nutrition and exercise, *Advances in Nutrition*; 2018 Nov 1; 9(6):813-23. [CrossRef], [Google Scholar], [Publisher]
- 41. Regoli F, Giuliani ME. Oxidative pathways of chemical toxicity and oxidative stress biomarkers in marine organisms, *Marine environmental research*; 2014 Feb 1; 93:106-17. [CrossRef], [Google Scholar], [Publisher]
- 42. Saras T. The Olive Oil Odyssey: Exploring the Richness of Nature's Liquid Gold, *Tiram Media*; 2023 Aug 11. [CrossRef], [Google Scholar], [Publisher]
- 43. Kaur C, Kapoor HC. Antioxidants in fruits and vegetables–the millennium's health, *International journal of food science & technology*; 2001 Oct 20; 36(7):703-25. [CrossRef], [Google Scholar], [Publisher]
- 44. Bansal AK, Bilaspuri GS. Impacts of oxidative stress and antioxidants on semen functions, *Veterinary medicine international*; 2011 Oct; 2011. [CrossRef], [Google Scholar], [Publisher]
- 45. Ferreira EM, Vireque AA, Adona PR, Meirelles FV, Ferriani RA, Navarro PA. Cytoplasmic

maturation of bovine oocytes: structural and biochemical modifications and acquisition of developmental competence, *Theriogenology*; 2009 Mar 15; 71(5):836-48. [CrossRef], [Google Scholar], [Publisher]

- 46. Kwiecien S, Jasnos K, Magierowski M, Sliwowski Z, Pajdo R, Brzozowski B, Mach T, Wojcik D, Brzozowski T. Lipid peroxidation, reactive oxygen species and antioxidative factors in the pathogenesis of gastric mucosal lesions and mechanism of protection against oxidative stress-induced gastric injury, J Physiol Pharmacol; 2014 Oct 1; 65(5):613-22. [CrossRef], [Google Scholar], [Publisher]
- 47. Bergamini CM, Gambetti S, Dondi A, Cervellati C. Oxygen, reactive oxygen species and tissue damage, *Current pharmaceutical design*; 2004 May 1; 10(14):1611-26. [CrossRef], [Google Scholar], [Publisher]
- 48. Kargapolova Y, Geißen S, Zheng R, Baldus S, Winkels H, Adam M. The enzymatic and nonenzymatic function of myeloperoxidase (MPO) in inflammatory communication, *Antioxidants*; 2021 Apr 5; 10(4):562. [CrossRef], [Google Scholar], [Publisher]
- 49. Iqbal Z, Sarkhosh A, Balal RM, Gómez C, Zubair M, Ilyas N, Khan N, Shahid MA. Silicon alleviate hypoxia stress by improving enzymatic and non-enzymatic antioxidants and regulating nutrient uptake in muscadine grape (Muscadinia rotundifolia Michx.), *Frontiers in Plant Science*; 2021 Feb 10; 11:2288. [CrossRef], [Google Scholar], [Publisher]
- 50. Hartman PE, Shankel DM. Antimutagens and anticarcinogens: a survey of putative interceptor molecules, *Environmental and Molecular Mutagenesis*; 1990; 15(3):145-82. [CrossRef], [Google Scholar], [Publisher]
- 51. Subedi P. Posttranslational Modification of 2-CysPrxs by Reduced Glutathione Interplays with CYP20-3-Dependent OPDA Signaling Pathways (Doctoral dissertation, Auburn

University). [CrossRef], [Google Scholar], [Publisher]

- 52. Karami S, Rahimi M, Babaei A. An overview on the antioxidant, anti-inflammatory, antimicrobial and anti-cancer activity of grape extract, *Biomed Res Clin Pract*; 2018; 3(2):1-4. [CrossRef], [Google Scholar], [Publisher]
- 53. Kaur C, Kapoor HC. Antioxidants in fruits and vegetables-the millennium's health, *International journal of food science & technology*; 2001 Oct 20; 36(7):703-25. [CrossRef], [Google Scholar], [Publisher]
- 54. Guven H, Arici A, Simsek O. Flavonoids in our foods: a short review, *Journal of Basic and Clinical Health Sciences*; 2019 Jan 6; 3(2):96-106. [CrossRef], [Google Scholar], [Publisher]
- 55. Stahl W, Sies H. Bioactivity and protective effects of natural carotenoids, *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*; 2005 May 30; 1740(2):101-7.
 [CrossRef], [Google Scholar], [Publisher]
- 56. Ciorîță A, Zăgrean-Tuza C, Moț AC, Carpa R, Parvu M. The phytochemical analysis of Vinca L. species leaf extracts is correlated with the antioxidant, antibacterial, and antitumor effects, *Molecules*; 2021 May 19; 26(10):3040. [CrossRef], [Google Scholar], [Publisher]
- 57. Abubakar AR, Haque M. Preparation of medicinal plants: Basic extraction and fractionation procedures for experimental purposes, *Journal of pharmacy & bioallied sciences*; 2020 Jan; 12(1):1. [CrossRef], [Google Scholar], [Publisher]
- 58. Jha AK, Sit N. Extraction of bioactive compounds from plant materials using combination of various novel methods: A review, *Trends in Food Science & Technology*; 2022 Jan 1; 119:579-91. [CrossRef], [Google Scholar], [Publisher]
- 59. Kamil Hussain M, Saquib M, Faheem Khan M. Techniques for extraction, isolation, and standardization of bio-active compounds from medicinal plants. Natural Bio-active Compounds: Volume 2: Chemistry,

Pharmacology and Health Care Practices. 2019:179-200. [CrossRef], [Google Scholar], [Publisher]

- 60. Gori A, Boucherle B, Rey A, Rome M, Fuzzati N, Peuchmaur M. Development of an innovative maceration technique to optimize extraction and phase partition of natural products, *Fitoterapia*; 2021 Jan 1; 148:104798. [CrossRef], [Google Scholar], [Publisher]
- 61. Siddiqui SA, Bahmid NA, Taha A, Khalifa I, Khan S, Rostamabadi H, Jafari SM. Recent advances in food applications of phenolicloaded micro/nanodelivery systems, *Critical Reviews in Food Science and Nutrition*; 2022 Mar 22:1-21. [CrossRef], [Google Scholar], [Publisher]
- 62. Doerr SH, Llewellyn CT, Douglas P, Morley CP, Mainwaring KA, Haskins C, Johnsey L, Ritsema CJ, Stagnitti F, Allinson G, Ferreira A. Extraction of compounds associated with water repellency in sandy soils of different origin, *Soil Research*; 2005 May 27; 43(3):225-37. [CrossRef], [Google Scholar], [Publisher]
- 63. Capote FP, De Castro ML. Analytical applications of ultrasound. Elsevier; 2007 Feb 14. [CrossRef], [Google Scholar], [Publisher]
- 64. Gulzar S. Lipids from Cephalothorax of Pacific White Shrimp (Litopenaeusvannamei) using Ultrasonic Assisted Extraction Process: Pretreatment, Enhanced Oxidative Stability and Applications (Doctoral dissertation, Prince of Songkla University). [CrossRef], [Google Scholar], [Publisher]
- 65. Todd R, Baroutian S. A techno-economic comparison of subcritical water, supercritical CO2 and organic solvent extraction of bioactives from grape marc, *Journal of Cleaner Production*; 2017 Aug 1; 158:349-58. [CrossRef], [Google Scholar], [Publisher]
- 66. Hemwimol S, Pavasant P, Shotipruk A. Ultrasound-assisted extraction of anthraquinones from roots of Morinda citrifolia, *Ultrasonics sonochemistry*; 2006 Sep

1; 13(6):543-8. [CrossRef], [Google Scholar], [Publisher]

- 67. Pateiro M, Gómez-Salazar JA, Jaime-Patlán M, Sosa-Morales ME, Lorenzo JM. Plant extracts obtained with green solvents as natural antioxidants in fresh meat products, *Antioxidants*; 2021 Jan 27; 10(2):181. [CrossRef], [Google Scholar], [Publisher]
- 68. Sultana B, Anwar F, Ashraf M. Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts, *Molecules*; 2009 Jun 15; 14(6):2167-80. [CrossRef], [Google Scholar], [Publisher]
- 69. Melgar-Lalanne G, Hernández-Álvarez AJ, Jiménez-Fernández M, Azuara E. Oleoresins from Capsicum spp.: Extraction methods and bioactivity, *Food and Bioprocess Tech*nology; 2017 Jan; 10:51-76. [CrossRef], [Google Scholar], [Publisher]
- 70. Mendonça JD, Guimarães RD, Zorgetto-Pinheiro VA, Fernandes CD, Marcelino G, Bogo D, Freitas KD, Hiane PA, de Pádua Melo ES, Vilela ML, Nascimento VA. Natural antioxidant evaluation: A review of detection methods, *Molecules*; 2022 Jun 1; 27(11):3563. [CrossRef], [Google Scholar], [Publisher]
- 71. Chua LY, Chong CH, Chua BL, Figiel A. Influence of drying methods on the antibacterial, antioxidant and essential oil volatile composition of herbs: A review, *Food and Bioprocess Technology*; 2019 Mar 15; 12:450-76. [CrossRef], [Google Scholar], [Publisher]
- 72. Grundtvig IP, Heintz S, Krühne U, Gernaey KV, Adlercreutz P, Hayler JD, Wells AS, Woodley JM. Screening of organic solvents for bioprocesses using aqueous-organic twophase systems, *Biotechnology advances*; 2018 Nov 15; 36(7):1801-14. [CrossRef], [Google Scholar], [Publisher]
- 73. Clark JH, Tavener SJ. Alternative solvents: shades of green, *Organic process research &*

development; 2007 Jan 19; 11(1):149-55. [CrossRef], [Google Scholar], [Publisher]

- 74. Ameer K, Shahbaz HM, Kwon JH. Green extraction methods for polyphenols from plant matrices and their byproducts: A review, Comprehensive Reviews in Food Science and Food Safety; 2017 Mar; 16(2):295-315. [CrossRef], [Google Scholar], [Publisher]
- 75. Gupta A, Naraniwal M, Kothari V. Modern extraction methods for preparation of bioactive plant extracts, *International journal* of applied and natural sciences; 2012 Aug; 1(1):8-26. [CrossRef], [Google Scholar], [Publisher]
- 76. de Fatima Alpendurada M. Solid-phase microextraction: a promising technique for sample preparation in environmental analysis, *Journal of Chromatography A*; 2000 Aug 11; 889(1-2):3-14. [CrossRef], [Google Scholar], [Publisher]
- 77. Périno-Issartier S, Abert-Vian M, Chemat F. Solvent free microwave-assisted extraction of antioxidants from sea buckthorn (Hippophae rhamnoides) food by-products, *Food and Bioprocess Technology*; 2011 Aug; 4:1020-8. [CrossRef], [Google Scholar], [Publisher]
- 78. Keke CO, Nwaogu LA, Igwe CU, Ekeke KL, Nsofor WN. Phytochemical and nutritional composition of ethanol-water leaf extracts of Justicia secunda and Jatropha tanjorensis, *GSC Biological and Pharmaceutical Sciences*; 2023; 23(3):042-53. [CrossRef], [Google Scholar], [Publisher]
- 79. Rahaiee S, Moini S, Hashemi M, Shojaosadati SA. Evaluation of antioxidant activities of bioactive compounds and various extracts obtained from saffron (Crocus sativus L.): a review, *Journal of Food Science and Technology*; 2015 Apr; 52:1881-8. [CrossRef], [Google Scholar], [Publisher]
- 80. Xu DP, Li Y, Meng X, Zhou T, Zhou Y, Zheng J, Zhang JJ, Li HB. Natural antioxidants in foods and medicinal plants: Extraction, assessment and resources, *International journal of*

molecular sciences; 2017 Jan 5; 18(1):96. [CrossRef], [Google Scholar], [Publisher]

- 81. Qayyum A, Sarfraz RA, Ashraf A, Adil S. Phenolic composition and biological (anti diabetic and antioxidant) activities of different solvent extracts of an endemic plant (Heliotropium strigosum), *Journal of the Chilean chemical society*; 2016 Mar; 61(1):2828-31. [CrossRef], [Google Scholar], [Publisher]
- 82. Mumtaz R, Zubair M, Khan MA, Muzammil S, Siddique MH. Extracts of Eucalyptus alba Promote diabetic wound healing by inhibiting α-glucosidase and stimulating cell proliferation, *Evidence-Based Complementary and Alternative Medicine*; 2022 Apr 15; 2022. [CrossRef], [Google Scholar], [Publisher]
- 83. Mokrani A, Madani K. Effect of solvent, time and temperature on the extraction of phenolic compounds and antioxidant capacity of peach (Prunus persica L.) fruit, *Separation and Purification Technology*; 2016 Apr 13; 162:68-76. [CrossRef], [Google Scholar], [Publisher]
- 84. Rajakaruna A, Manful CF, Abu-Reidah IM, Critch AL, Vidal NP, Pham TH, Cheema M, Thomas R. Application of solvent pH under pressurized conditions using accelerated solvent extraction and green solvents to extract phytonutrients from wild berries, *Food Bioscience*; 2022 Jun 1; 47:101471. [CrossRef], [Google Scholar], [Publisher]
- 85. Cano A, Arnao MB. ABTS/TEAC (2, 2'-azinobis (3-ethylbenzothiazoline-6-sulfonic acid)/Trolox®-Equivalent Antioxidant Capacity) radical scavenging mixed-mode assay, *Measurement of antioxidant activity & capacity: recent trends and applications*; 2018 Jan 11; 117-39. [CrossRef], [Google Scholar], [Publisher]
- 86. Kumar BR. Application of HPLC and ESI-MS techniques in the analysis of phenolic acids and flavonoids from green leafy vegetables (GLVs), *Journal of pharmaceutical analysis*;

2017 Dec 1; 7(6):349-64. [CrossRef], [Google Scholar], [Publisher]

- 87. Moon JK, Shibamoto T. Antioxidant assays for plant and food components, *Journal of agricultural and food chemistry*; 2009 Mar 11; 57(5):1655-66. [CrossRef], [Google Scholar], [Publisher]
- 88. Wu H, Guo J, Chen S, Liu X, Zhou Y, Zhang X, Xu X. Recent developments in qualitative and quantitative analysis of phytochemical constituents and their metabolites using liquid chromatography–mass spectrometry, *Journal of pharmaceutical and biomedical analysis*; 2013 Jan 18; 72:267-91. [CrossRef], [Google Scholar], [Publisher]
- 89. Iloki-Assanga SB, Lewis-Luján LM, Lara-Espinoza CL, Gil-Salido AA, Fernandez-Angulo D, Rubio-Pino JL, Haines DD. Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of Bucida buceras L. and Phoradendron californicum, *BMC research notes*; 2015 Dec; 8(1):1-4. [CrossRef], [Google Scholar], [Publisher]
- 90. Zhou X, Li CG, Chang D, Bensoussan A. Current status and major challenges to the safety and efficacy presented by Chinese herbal medicine, *Medicines*; 2019 Jan 18; 6(1):14. [CrossRef], [Google Scholar], [Publisher]
- 91. Marino A, Battaglini M, Moles N, Ciofani G. Natural antioxidant compounds as potential pharmaceutical tools against neurodegenerative diseases, *ACS omega*; 2022 Jul 19; 7(30):25974-90. [CrossRef], [Google Scholar], [Publisher]
- 92. Gonzalez E, Vaillant F, Pérez AM, Rojas G. In vitro cell-mediated antioxidant protection of human erythrocytes by some common tropical fruits. [CrossRef], [Google Scholar], [Publisher]
- 93. Gonzalez E, Vaillant F, Pérez AM, Rojas G. In vitro cell-mediated antioxidant protection of human erythrocytes by some common

tropical fruits. [CrossRef], [Google Scholar], [Publisher]

- 94. Chawla R, Thakur P, Chowdhry A, Jaiswal S, Sharma A, Goel R, Sharma J, Priyadarshi SS, Kumar V, Sharma RK, Arora R. Evidence based herbal drug standardization approach in coping with challenges of holistic management of diabetes: a dreadful lifestyle disorder of 21st century, *Journal of Diabetes & Metabolic Disorders*; 2013 Dec; 12:1-6. [CrossRef], [Google Scholar], [Publisher]
- 95. Cabrera C, Giménez R, López MC. Determination of tea components with antioxidant activity, *Journal of agricultural and food chemistry*; 2003 Jul 16; 51(15):4427-35. [CrossRef], [Google Scholar], [Publisher]
- 96. Chang J. Medicinal herbs: drugs or dietary supplements?, *Biochemical pharmacology*; 2000 Feb 1; 59(3):211-9. [CrossRef], [Google Scholar], [Publisher]
- 97. Ushie O.A., Bako B., Malu S. P. and Danladi A. H. (2023). Phytochemistry and Antioxidants Activities of Four Different Solvent Extracts of *Justicia secunda* Stem. FUAM Journal of Pure and Applied Science, 3(1):59-68. [CrossRef], [Google Scholar], [Publisher]
- 98. Bako, B., Danladi, A. H., Bulus, G. G., & Ushie, O. A. (2023). Quantitative analysis of selected phytochemicals and antimicrobial potentials in *Justicia secunda* leaf crude extracts. WJPMR, 9(8), 228-232. [CrossRef], [Google Scholar], [Publisher]
- 99. Bako, B., Ushie, O. A., Malu, S. P., & Kendeson, A. C. (2023). Phytochemical screening and antioxidant activities of four different solvent extracts of Justicia secunda leaf extracts. *Journal of Chemical Research Advances*, 4(1), 15-20. Phytochemical screening and antioxidants activities of four different solvent extracts of Justicia secunda leaf extracts - JCRA - Journal of Chemical Research Advances. [CrossRef], [Google Scholar], [Publisher]
- 100. Osioma E, Hamilton-Amachree A. Comparative study on the phytochemical and

in vitro antioxidant properties of methanolic leaf extract of Justicia secunda Vahl, *Nigerian Journal of Science and Environment*; 2017; 15(1):111-7. [CrossRef], [Google Scholar], [Publisher]

- 101. Li H, Zhang D, Tan LH, Yu B, Zhao SP, Cao WG. Comparison of the antioxidant properties of various solvent extracts from Dipsacus asperoides and identification of phenolic compounds by LC-ESI-QTOF-MS–MS, *South African Journal of Botany*; 2017 Mar 1; 109:1-8. [CrossRef], [Google Scholar], [Publisher]
- 102. Bako, B., Ushie, O. A., & Malu, S. P. (2023). Lupeol and Lauric Acid Isolated from Ethyl Acetate Stem Extract of Justicia Secunda and their Antimicrobial Activity. *Journal of Chemical Society of Nigeria*, 48(1). [CrossRef], [Google Scholar], [Publisher]
- 103. Bulus Bako. Applications of Justicia secunda Extracts in Functional Foods and Natural Products: A Review, Adv. J. Chem. B, 6 (2024)1-10[CrossRef], [Google Scholar], [Publisher]